

High Inorganic Carbon Extraction Capacity of Submerged Macrophytes from Soft-water Faroese Lakes

Vatnplantur í føroyskum vøtnum við bleytum vatni taka lættliga ólívrannið kolevni upp

Tom Vindbæk Madsen and Jens Bagger

Department of Plant Ecology, University of Aarhus, Nordlandsvej 68, DK-8240 Risskov, Denmark

Corresponding author: Tom Vindbæk Madsen, E-mail: Tom.Madsen@biology.au.dk

Úrtak

Vatnplantur (macrophytes) úr tíggju føroyskum vøtnum við millum 0.16 og 0.8 meqv. L⁻¹ av alkali í sær vóru tiknar, og kannað varð, hvussu væl tær vóru førar fyri at taka ólívrannið kolevni upp. Av 26 sløgum, ið vórðu kannað, tók um leið helmingurin væl av kolevni upp samsvarandi við, at tær vóru førar fyri at miðsavna kolevni. Hin helmingurin sýndi eyðkenni samsvarandi við spjadda CO₂ veiting og C-3 fotosamanrenning. Hóast lítið alkali var í føroyskum vøtnum og sostatt lítið hydrokarbonati, megnaðu tey sløg, sum eru før fyri at taka kolevni upp, at troyta tað hydrokarbonat, sum var. Einkin samband var funnið millum alkali í vøtnunum og førleikan hjá plantunum at taka upp kolevni.

Abstract

Submerged aquatic macrophytes from ten Faroese lakes with an alkalinity between 0.16 and 0.8 meqv L⁻¹ were collected and their inorganic carbon extraction capacity determined. Among the 26 species tested, about one-half had a high carbon extraction capacity consistent with possession of a carbon concentrating mechanism. The other one-half showed characteristics consistent with diffusive CO₂ supply and C-3 photosynthesis. Despite the low alkalinity of the Faroese lakes and, thus, low bicarbonate concentration, the species employing a

carbon concentrating mechanism were able to exploit the bicarbonate pool of the water. No relationship between lake alkalinity and carbon extraction capacity of the plants was observed.

Introduction

Submerged plants in lakes and streams are often exposed to environmental conditions significantly different from conditions existing in terrestrial habitats from which vascular plants are believed to have colonised freshwater habitats (Raven, 1995). In particular, the supply rate of CO₂ and O₂ are substantially lower in water than in air due to the 10⁴ times lower diffusion rate of gases. The low diffusion rate impedes the gas flux across the stagnant layer of water surrounding the plants and, thereby, restricts the exchange rate between plants and bulk water. As a result, CO₂ saturation for submerged macrophyte photosynthesis is reached only at relatively high concentra-

tions, often above 500 μM (Steeman-Nielsen, 1947; Maberly and Spence, 1989; Nielsen and Sand-Jensen, 1993; Larcher, 1995). This is more than 30 times the equilibrium concentration in water, which is about 16 μM (at 15°C). However, due to metabolic activity and a slow exchange rate between water and air, disequilibrium is often encountered. Thus, in a survey comprising 1835 lakes and 4665 samples (Cole *et al.* (1994) found that nearly all had CO_2 concentrations different from air equilibrium, with 87% having higher concentrations.

The high CO_2 requirement for saturation of photosynthesis by submerged macrophytes in relation to the low concentrations found in most waters suggests that CO_2 limitation of photosynthesis may be a common phenomenon, an hypothesis that has been confirmed in several investigations (Maberly, 1985; Madsen and Maberly, 1991; Madsen, 1991; Nielsen and Sand-Jensen, 1993; Frost-Christensen and Sand-Jensen, 1995). The CO_2 limitation of photosynthesis might be circumvented or ameliorated by the plants through various structural and physiological adaptations. These mechanisms and strategies can be grouped into two main categories: spatial escape and biochemical/physiological adaptations. Plants that rely on spatial escape grow in habitats rich in CO_2 , have the ability to develop aerial leaves or can use sediment CO_2 . Biochemical and physiological adaptations include bicarbonate use and different types of C-4 photosynthesis. The biochemical/physiological adaptations are often referred to as Carbon Concentrating

Mechanisms (CCM), due to their capacity to enhance the internal concentration above what can be obtained by passive CO_2 uptake and C-3 photosynthesis.

Escape Strategies

In the bulk water of lakes, the concentration of CO_2 might be higher than atmospheric equilibrium concentrations, though concentrations sufficient to saturate photosynthesis of submerged macrophytes are rare (Cole *et al.*, 1994). Higher CO_2 concentrations can also be encountered immediately adjacent to the surface of organic sediment with high bacterial activity (Pedersen *et al.*, 1995). An inorganic carbon source that might be exploited by low stature plants growing close to the sediment.

A quantitatively significant use of sediment- CO_2 among freshwater macrophytes is a widespread trait among the isoetids of which *Littorella uniflora*, *Lobelia dortmanna* and *Isoetes lacustris* are prominent members (Wium-Andersen, 1971; Boston *et al.*, 1987). The isoetids are small plants with short, stiff rosette leaves, a high root: shoot ratio and a well-developed lacunal system. The lacunal system forms a more or less continuous airspace along the entire length of the plants through which CO_2 diffuses from the sediment via the roots to the leaves and oxygen diffuses in the opposite direction (Sculthorpe, 1967). The ecological advantage of sediment- CO_2 use lies in the high CO_2 concentration in the interstitial water, which may be 50 to 100 times higher than in the bulk water (Wium-Andersen and Andersen, 1972; Roelofs *et al.*, 1984; Boston *et al.*, 1987). Despite the high

CO₂ concentration in the interstitial water, the dependence on diffusive transport sets a limit on the maximum leaf-length that can be obtained by the isoetid as diffusion is slow over longer distances even in air. Consequently, the isoetids often dominate the flora of oligotrophic, soft-water lakes, but cannot compete with the taller elodeid vegetation in more alkaline systems.

Use of the more readily accessible atmospheric CO₂ is a characteristic of amphibious plants that have the ability to grow submerged as well as emergent. The exploitation of atmospheric CO₂ requires that the plants develop aerial leaves, either floating on or raised above the water surface. The aerial leaves have functional stomata and resemble leaves of terrestrial plants by being thicker and having a higher photosynthetic capacity and CO₂ affinity than their submerged counterparts when measured in air (Sculthorpe, 1967; Sand-Jensen and Frost-Christensen, 1999).

Biochemical and Physiological Adaptations

A number of aquatic macrophyte species have CO₂ uptake characteristics that deviate from those expected for plants relying on diffusive entry of CO₂ with subsequent fixation by the C-3 carbon fixation pathway. In particular, the rate of photorespiration and the CO₂ compensation point are lower than predicted. This indicates that these plants possess a carbon acquisition or concentrating system that can overcome the limitation imposed by CO₂ diffusion and C-3 photosynthesis. Among these systems, bicarbonate use is the most widespread (Madsen and Sand-Jensen, 1991),

whereas C-4 photosynthesis and Crassulacean Acid Metabolism (CAM) is employed by only a few species.

In addition to CO₂, inorganic carbon in water exists as bicarbonate (HCO₃⁻) and carbonate (CO₃²⁻) and the three carbon species form a buffering system, where the relative distribution among species is determined by pH and, to a lesser extent, by temperature and salinity (Stumm and Morgan, 1996). With increasing pH, the equilibrium shifts from CO₂ to bicarbonate and carbonate with pK_a values of 6.38 and 10.38 (20°C) (Prins and Elzenga, 1989). Thus, in waters with pH above 6.4, the concentration of HCO₃⁻ is higher than the CO₂ concentration. In alkaline waters, plant species with the ability to use bicarbonate, therefore, have an advantage compared to plants that use CO₂ only, an advantage that is greater at higher alkalinity and pH. Based on characteristic electro-chemical potentials of plant cells, it can be calculated that passive entry of HCO₃⁻ only takes place if the external HCO₃⁻ concentrations is 10²–10⁴ times higher than internal concentrations, an observation that has led to the conclusion that HCO₃⁻ uptake is active (Raven, 1970).

Among freshwater species possessing the ability to use bicarbonate, the affinity for HCO₃⁻ in general is much lower than the affinity for CO₂, when expressed as the concentration needed to half-saturate photosynthesis and as the initial slope of the dose-response curve (Madsen and Sand-Jensen, 1991; Madsen, 1993; Maberly and Madsen, 1998). The lower affinity for bicarbonate compared to CO₂ potentially re-

duces the ecological significance of HCO_3^- use, in particular in soft-water and acid lakes and streams.

C-4 photosynthesis has so far been found in *Hydrilla verticillata* only (Reiskind *et al.*, 1997). *Hydrilla* lacks Kranz anatomy to separate the two carboxylation systems of C-4, as known from terrestrial C-4 plants. Instead, the two systems appear to be separated at the cell level, as revealed by immunocytochemical gold-labelling and fluorescence techniques (Reiskind *et al.*, 1989). C-4 carboxylation and phosphoenolpyruvate carboxylase (PEPcase) is found in the cytosol and C-3 carboxylation and ribulose biphosphate carboxylase-oxygenase is confined to the chloroplasts. The ecophysiological benefit of C-4 is the suppression of photorespiration, through which 50% of the fixed carbon may be lost as respiratory CO_2 in C-3 plants.

A number of submerged macrophytes, most belonging to the isoetids (Keeley, 1996), have developed an inorganic carbon uptake system that resembles the terrestrial CAM plant system and allows the plants to take up inorganic carbon throughout most of the light/dark cycle ((Madsen, 1987a). The main benefit of CAM for isoetids in soft-water lakes with low inorganic carbon concentrations is by far the extended period available for carbon uptake. However, it appears that CAM may also have a stimulating effect on net photosynthesis, due to an inhibitory effect on photorespiration caused by the high internal CO_2 concentrations encountered during decarboxylation (Madsen, 1987a).

The physiological and biochemical

adaptations in submerged macrophytes as described above are believed to be directed towards maximising inorganic carbon uptake efficiency. The trade-off is enhanced running costs for inorganic carbon uptake and the need for investments in uptake systems. The ability to use CCM is not a constant feature, however, but appears to be regulated in response to growth conditions and only shows high activity when needed, *i.e.* when the availability of inorganic carbon is low relative to the demand. Thus, bicarbonate-users often down-regulate their affinity and capacity for HCO_3^- uptake when grown at high CO_2 concentration (Sand-Jensen and Gordon, 1993). Similarly, CAM activity has been found to be suppressed for plants growing at high CO_2 or in low light, where the inorganic carbon requirement is low (Madsen, 1987b).

In the present study, we measured the carbon extraction capacity for a range of submerged macrophyte species from lakes of the Faroe Islands. The lakes are all soft-water and low in inorganic carbon (H.-H. Schierup, pers. comm.) and support a submerged macrophyte flora that includes numerous species known from the literature to have the potential for using bicarbonate in photosynthesis (Hansen, 1966; Maberly and Spence, 1983). The specific objectives of the study were 1) to examine if species known to be bicarbonate users in alkaline lakes maintain the ability in the more soft-water, Faroese lakes, where the low affinity for HCO_3^- may restrict the ecological advantage of HCO_3^- use, and 2) to test whether the extraction capacity of the plants was related to lake water alkalinity.

Table 1. Alkalinity (mean \pm s.d, $n = 2-3$) and UTM co-ordinates (zone 29V) for lakes included in this investigation.

Talva 1. Alkani (Miðal \pm s.d., $n=2-3$) og UTM samskiparar (øki 29 V) yvir vøtn, sum henda kanning fevnir um.

Lake	UTM co-ordinates	Alkalinity (meqv L ⁻¹)
Eysturoy		
Toftavatn	6897580 m.N, 619070 m.E	0.16 \pm 0.016
Múlavík	6909700 m.N, 599250 m.E	0.17 \pm 0.019
Streymoy:		
Saksunarvatn	6902600 m.N, 595940 m.E	0.19 \pm 0.013
Mjáuvøtn	6889940 m.N, 604700 m.E	0.13 \pm 0.016
Áarstiggjafossur	6861550 m.N, 605760 m.E	0.29 \pm 0.005
Pond at Tórshavn	6878290 m.N, 616490 m.E	0.81 \pm 0.058
Sandoy:		
Norðara Hálsavatn	6863700 m.N, 612100 m.E	0.18 \pm 0.025
Sandsvatn	6859600 m.N, 614900 m.E	0.37 \pm 0.044
Gróthúsvatn	6858620 m.N, 614220 m.E	0.37 \pm 0.021
Stórvatn	6858400 m.N, 609140 m.E	0.31 \pm 0.016

Material and methods

Specimens of 26 aquatic macrophyte species and water were collected in ten lakes on Eysturoy, Streymoy and Sandoy (Table 1). Several of the species were collected in more than one lake.

Alkalinity of the lake water was measured by Gran titration (Mackereth *et al.*, 1989) of 20 ml samples with 0.1 M HCl using a Radiometer PHM 84 pH-meter with a glass combination electrode (Radiometer GK2401C).

The carbon extraction capacity of the plants was measured employing pH drift experiments. Small apical shoots, young, but with fully developed leaves or thallus segments, were incubated in closed, 25 ml glass bottles and allowed to photosynthesise for 18 hours. The incubation medium was modified from Smart and Barko (1985) by adjusting the amount of KHCO₃ and NaHCO₃ to give alkalinities of 0.125 meqv L⁻¹ and 1.0 meqv L⁻¹. The medium was

bubbled with atmospheric air prior to incubation. The bottles were placed on a rotating wheel in a water-bath at 10°C \pm 0.1°C and light was provided at 250 μ mol photons m⁻² s⁻¹ by a high-pressure metal halide lamp. After 16 hours of incubation, pH was measured and alkalinity determined for 10 ml samples by Gran titration. The carbon extraction capacity is expressed as final-pH.

The concentration of the different inorganic carbon species was calculated from alkalinity, pH and temperature, as described in Maberly (1996).

Results and discussion

The selection of species included in this study is listed in Table 2.

The final-pH reached in the pH drift experiments at an alkalinity of 1.0 meqv L⁻¹ varied among the different plant species from 8.55 for *Isoetes lacustris* to 10.87 for the filamentous algae *Vaucheria* sp. (Table

Table 2.

List of species included in this study. Alternative inorganic carbon sources that can be exploited by the different vascular plants and macroalgae are listed. Ability to use sediment-CO₂ was extracted from Spence and Maberly (1985) and Richardson et al. (1984) and the ability to exploit the atmospheric CO₂ pool was diagnosed from the ability to develop floating or aerial leaves.

Talva 2

Yvirlit yvir slögini, sum henda kanning fevnir um. Listi yvir aðrar ólífurrunnar kolevniskeldur, sum tær ymisku leggstreingjaplanturnar og makroalgurnar fáa nýtt. Spence og Maberly (1985) og Richardson et al. (1984) komu fram á, hvussu væl planturnar vóru førar fyri at nýta CO₂ úr botnsetingini, og alt eftir, hvussu tær megnaðu at menna flóti-ella luftbløð, varð mettt um, hvussu væl tær vóru førar fyri at nýta CO₂-puljuna í luftini.

Species	Alternative Carbon Source
<i>Callitriche stagnalis</i>	Atmosphere
<i>C. hamulata</i>	Atmosphere
<i>Eleocharis acicularis</i>	Atmosphere
<i>Fontinalis antipyretica</i>	
<i>Isoetes lacustris</i>	Sediment
<i>Juncus bulbosus</i>	
<i>Littorella uniflora</i>	Sediment
<i>Lobelia dortmanna</i>	Sediment
<i>Myriophyllum alterniflorum</i>	
<i>Nymphaea alba</i>	Atmosphere
<i>Polygonum amphibium</i>	Atmosphere
<i>Potamogeton alpinus</i>	Atmosphere
<i>P. filiformis</i>	
<i>P. gramineus</i>	Atmosphere
<i>P. natans</i>	Atmosphere
<i>P. obtusifolius</i>	
<i>P. perfoliatum</i>	
<i>P. polygonifolius</i>	Atmosphere
<i>P. praelongus</i>	
<i>Sparganium angustifolium</i>	Atmosphere
<i>Utricularia vulgaris</i>	
<i>Chara</i> sp.	
<i>Cladophora</i> sp.	
<i>Nitella</i> sp.	
<i>Spirogyra</i> sp.	
<i>Vaucheria</i> sp.	

3). Calculated and expressed as the percentage of the inorganic carbon pool extracted from the water, *Isoetes* had removed less than 1%, whereas *Vaucheria* had removed more than 57%.

The differences in extraction capacity among plants are believed to reflect differences in their inorganic carbon uptake mechanisms. For species with low extraction capacity, the data are consistent with diffusive entry of CO₂ and C-3 physiology. In contrast, species with high extraction capacity are expected to employ some kind of carbon concentrating mechanism. Of these mechanisms, active uptake of bicarbonate constitutes the most efficient system in terms of inorganic carbon extraction capacity among submerged macrophytes (Madsen and Sand-Jensen, 1991). Whether plants rely on diffusive entry of CO₂ and C-3 photosynthesis or use a carbon concentrating system cannot be determined from final-pH alone. However, from final-pH and alkalinity of the incubation medium, the CO₂ compensation point of the plants can be calculated and compared to CO₂ compensation points reported in the literature, which vary from about 1.5 μ M to more than 100 μ M (Maberly and Spence, 1983). For plants incubated in a medium with an alkalinity of 1.0 meqv L⁻¹ at 10 °C, as in this study, a compensation point of 1.5 μ M will allow the plants to raise pH to about 9.2. Using this value to separate species with and without the ability to operate a CCM, the data in Table 3 show that 14 of the 26 species tested had this ability. Table 3 also shows, however, that the pH drift data do not offer a marked distinction be-

tween non-CCM and CCM species. Rather, the species show a continuous gradient in carbon extraction capacity, which reflects genotypical differences in carbon uptake efficiency and the great plasticity in inorganic carbon uptake physiology generally observed for aquatic macrophytes. For species restricted to CO₂ use and for species running a CCM, the affinity for inorganic carbon is greatly dependent on environmental conditions. Under conditions where the restriction on inorganic carbon uptake is high, the plants respond by enhancing the resource allocation towards carbon uptake, resulting in an improved affinity and extraction capacity for inorganic carbon (Sand-Jensen and Gordon, 1993; Madsen, 1993; Madsen *et al.*, 1996). The restriction of inorganic carbon uptake rates is not governed by the supply rate of CO₂ and other inorganic carbon species alone; of more importance is the balance between the supply rate and the demand for inorganic carbon by the plants. Environmental parameters other than inorganic carbon will, therefore, affect the extent of inorganic carbon limitation and the extent of limitation will be inversely coupled to the extent to which other factors limit plant growth and, thereby, affect inorganic carbon demand. Thus, it cannot be excluded that among species in the border-zone between non-CCM and CCM species some species categorised as non-CCM might actually have a concentrating system with a very low efficiency and vice versa.

A number of the species tested and found to rely on diffusive entry of CO₂ and C-3 photosynthesis have developed mecha-

Table 3.

Inorganic carbon extraction capacity for submerged macrophyte species in Faroese lakes. Inorganic carbon extraction capacity was measured by pH-drift experiments at an alkalinity of 1.0 and 0.125 meqv L⁻¹ and is expressed as final-pH after 16 hours incubation at 10 fC and 250 (mol m⁻² s⁻¹ PAR. Mean (s.d., n = 3-6; n.a. = not analysed.

Talva 3.

Førleiki hjá vatnplantusløgum í føroyskum vøtnum at taka ólívrinnið kolevni upp. pH-rekroyndir mátaðu førleikan at taka ólívrinnið kolevni upp, tá ið alkalið er 1.0 og 0.125 meqv. L⁻¹ og verður nevnt endaligt pH, tá ið tilgerðartíðin er 16 tímar við 10 °C og 250 (mol m⁻² s⁻¹ PAR. Miðal (s.d.) n=3-6; n.a.=ikki sundurgreinað.)

Species	Final-pH	
	1.0 meqv L ⁻¹	0.125 meqv L ⁻¹
<i>Nymphaea alba</i>	8.44±0.16	n.a.
<i>Isoetes lacustris</i>	8.55±0.04	n.a.
<i>Sparganium angustifolium</i>	8.58±0.04	n.a.
<i>Lobelia dortmanna</i>	8.62±0.25	n.a.
<i>Callitriche stagnalis</i>	8.72±0.03	n.a.
<i>Littorella uniflora</i>	8.75±0.24	n.a.
<i>Eleocharis acicularis</i>	8.93±0.15	n.a.
<i>Fontinalis antipyretica</i>	9.02±0.01	n.a.
<i>Juncus bulbosus</i>	9.07±0.02	n.a.
<i>Nitella</i> sp.	9.07±0.05	n.a.
<i>Potamogeton polygonifolius</i>	9.19±0.06	n.a.
<i>Callitriche hamulata</i>	9.27±0.09	n.a.
<i>Potamogeton natans</i>	9.52±0.47	8.52±0.56
<i>Utricularia vulgaris</i>	9.70±0.18	9.43±0.52
<i>Potamogeton alpinus</i>	10.00±0.10	9.92±0.08
<i>P. praelongus</i>	10.03±0.08	9.96±0.07
<i>P. gramineus</i>	10.12±0.16	9.85±0.10
<i>Myriophyllum alterniflorum</i>	10.13±0.02	9.82±0.08
<i>Potamogeton obtusifolius</i>	10.21±0.07	9.51±0.23
<i>P. filiformis</i>	10.26±0.09	9.64±0.03
<i>Spirogyra</i> sp.	10.39±0.41	9.80±0.42
<i>Potamogeton perfoliatus</i>	10.44±0.04	10.22±0.24
<i>Chara</i> sp.	10.52±0.02	9.62±0.07
<i>Cladophora</i> sp.	10.74±0.10	10.26±0.07
<i>Vaucheria</i> sp.	10.87±0.16	9.01±0.59

nisms that allow exploitation of inorganic carbon sources other than those in the bulk water (Table 2). The isoetids (*Littorella uniflora*, *Isoetes lacustris* and *Lobelia dortmanna*) use sediment- CO_2 . In addition, *Littorella* and *Lobelia*, including the Faroese specimen, are CAM plants (Keeley, 1996; Madsen and Bagger, unpubl. results 1999) and, thereby, have the potential to improve the carbon balance by allowing inorganic carbon uptake in both light and dark. However, the effects of CAM on the carbon extraction capacity appear to be low (Table 3). Of the remaining non-CCM species, *Sparganium angustifolium*, *Potamogeton polygonifolius* and the two *Callitriche* species have the ability to develop aerial or floating leaves allowing acquisition of CO_2 from the more readily available atmospheric pool. *Eleocharis acicularis*, *Juncus bulbosus*, *Nitella* sp. and *Fontinalis antipyretica* are not known to have developed traits or features improving inorganic carbon uptake (Steeman-Nielsen, 1947; Bain and Proctor, 1980; Allen and Spence, 1981; Morton and Keeley, 1990), except for one report showing that *Fontinalis antipyretica* collected in a Spanish river could use bicarbonate ((Penuelas, 1985). These species, however, often grow close to the sediment and might benefit from the higher CO_2 concentration found there (Maberly, 1985).

The list of species with CCM (final-pH > 9.5, Table 3) agrees with reports in the literature (Spence and Maberly, 1985), except for submerged leaves of *Potamogeton natans* that have been reported to be unable to operate a CCM (Maberly and Spence, 1983). The CO_2 compensation point calcu-

lated for *P. natans* varied among sites, but was very low (0.2 μM to 0.6 μM) and unlikely to be achievable without a carbon concentrating system. Interference from epiphytes cannot be ruled out, however, although no visible biofilm was found. The CO_2 compensation point reported for *P. natans* by Maberly and Spence (1983) for submerged leaves was 2.1 μM , which is comparable to the range reported by Frost-Christensen and Sand-Jensen (1995) for floating leaves measured in water. For *Utricularia vulgaris* no report on CCM has been found. Another *Utricularia* species, *U. purpurea*, has been reported to be unable to use bicarbonate, though tested at low alkalinity (<0.07 meqv L^{-1}) only (Moeller, 1978). In contrast, *Utricularia australis* has a high carbon extraction capacity (L. Kristiansen, pers. comm.).

For most of the species categorised as having CCM, only three, *P. natans*, *P. alpinus* and *P. gramineus*, have the ability to explore alternative inorganic carbon sources, all by the ability to develop floating leaves.

Though not verified directly, it is assumed that the CCM operating in the species tested was based on HCO_3^- use, which is the most widespread among freshwater macrophytes (Prins and Elzenga, 1989; Madsen and Sand-Jensen, 1991). For these species, the HCO_3^- compensation point, calculated from alkalinity and final-pH, varied from 0.1 to 0.77 mM. This range is comparable to the range reported for other macrophytes measured at similar alkalinity (0.5 – 2.0 mM), but substantially higher than the compensation point found

for the more efficient microalgae (Allen and Spence, 1981).

The alkalinity of the Faroese lakes from which the plants were collected varied between 0.16 and 0.37 meqv L⁻¹ (Table 1) and the concentration of bicarbonate between 0.16 and 0.37 mM (except for one lake with an alkalinity of 0.8 meqv L⁻¹). These bicarbonate concentrations are lower than the compensation point of most of the tested species (0.37 mM HCO₃⁻ is equivalent to a final-pH of about 10.3). To test whether the compensation point was affected by alkalinity, pH drift experiments were performed at 0.125 meqv L⁻¹ for species found to have a CCM.

The final-pH at 0.125 meqv L⁻¹ was lower than at 1 meqv L⁻¹ (Table 3), which is a result of the lower, total inorganic carbon concentration at low, compared to high, alkalinity at a given pH. The calculated HCO₃⁻ compensation points, which ranged from 0.12 to 0.03 mM, were also lower at 0.125 meqv L⁻¹ than at 1.0 meqv L⁻¹. The lower compensation points show that the CCM species will be able to use HCO₃⁻ even at the low concentrations found in the Faroese lakes. The relative contribution of HCO₃⁻ to total inorganic carbon uptake cannot, however, be estimated from pH-drift experiments alone.

The inorganic carbon uptake characteristics of aquatic macrophytes are very plastic within a particular species and change in response to the availability of inorganic carbon and other resources. For *Elodea canadensis* collected in a range of Danish lakes and streams the affinity for bicarbonate has been found to vary from high to no

measurable affinity in response to differences in bicarbonate and CO₂ concentrations in the water (Sand-Jensen and Gordon, 1993). Also, for plants grown in the laboratory, substantial acclimation in affinity for both HCO₃⁻ and CO₂ and in carbon extraction capacity was observed, not only in response to CO₂ and HCO₃⁻ availability (Madsen *et al.*, 1996), but also in response to other environmental parameters (Madsen, unpubl. results, 1998). For the species collected in this study, however, no correlation was observed between carbon extraction capacity among species and alkalinity of water from the collection site, nor was the distribution of species related to lake alkalinity. These results likely reflect the narrow range in alkalinity found in the Faroese lakes sampled rather than the lack of physiological plasticity and responsiveness among the species.

Acknowledgement

Funding for this study was provided by The Danish Research Agency, Grant No. 9803000.

References

- Allen, E.D. and Spence, D.H.N. 1981. The differential ability of aquatic plants to utilize the inorganic carbon supply in fresh waters. *New Phytol.* 87: 269-283.
- Bain, J.T. and Proctor, M.C.F. 1980. The requirement of aquatic bryophytes for free CO₂ as an inorganic carbon source: Some experimental evidence. *New Phytol.* 86: 393-400.
- Boston, H.L., Adams, M.S. and Pienkowski, T.P. 1987. Utilization of sediment CO₂ by selected North American isoetids. *Ann. Bot.* 60: 485-494.
- Cole, J.J., Caraco, N.F., Kling, G.W. and Kratz, T.K. 1994. Carbon dioxide supersaturation in the surface waters of lakes. *Science* 265: 1568-1570.
- Frost-Christensen, H. and Sand-Jensen, K. 1995. Comparative kinetics of photosynthesis in floating and

- submerged *Potamogeton* leaves. *Aquat. Bot.* 51: 121-134.
- Hansen, K. 1966. Vascular plants in the Faeroes. Horizontal and vertical distribution. *Dansk Bot. Arkiv* 24: 7-141.
- Keeley, J.E. 1996. Aquatic CAM photosynthesis. In: Winter, K., Smith, J.A.C. (eds.). *Crassulacean Acid Metabolism. Biochemistry, Ecophysiology and Evolution*. 281-295.
- Larcher, W. 1995. (3rd ed). *Physiological Plant Ecology*. Berlin.
- Maberly, S.C. 1985. Photosynthesis by *Fontinalis antipyretica*. I. Interaction between photon irradiance concentration of carbon dioxide and temperature. *New Phytol.* 100: 127-140.
- Maberly, S.C. 1996. Diel, episodic and seasonal changes in pH and concentrations of inorganic carbon in a productive lake. *Freshw. Biol.* 35: 579-598.
- Maberly, S.C. and Madsen, T.V. 1998. Affinity for CO₂ in relation to the ability of freshwater macrophytes to use HCO₃⁻. *Func. Ecol.* 12: 99-106.
- Maberly, S.C. and Spence, D.H.N. 1983. Photosynthetic inorganic carbon use by freshwater plants. *J. Ecol.* 71: 705-724.
- Maberly, S.C. and Spence, D.H.N. 1989. Photosynthesis and photorespiration in freshwater organisms: amphibious plants. *Aquat. Bot.* 34: 267-286.
- Mackereth, F.J.H., Heron, J. and Talling, J.F. 1989. *Water Analysis: Some revised methods for limnologists*. Kendal.
- Madsen, T.V. 1987a. Interactions between internal and external CO₂ pools in the photosynthesis of the aquatic CAM plants *Littorella uniflora* (L.) Aschers. and *Isoetes lacustris* L. *New Phytol.* 106: 35-50.
- Madsen, T.V. 1987b. The effect of different growth conditions on dark and light carbon assimilation in *Littorella uniflora*. *Physiol. Plant.* 70: 183-188.
- Madsen, T.V. 1991. Inorganic carbon uptake kinetics of the stream macrophyte *Callitriche cophocarpa* Sendt. *Aquat. Bot.* 40: 321-332.
- Madsen, T.V. 1993. Growth and photosynthetic acclimation by *Ranunculus aquatilis* L. in response to inorganic carbon availability. *New Phytol.* 125: 707-715.
- Madsen, T.V. and Maberly, S.C. 1991. Diurnal variation in light and carbon limitation of photosynthesis by two species of submerged freshwater macrophyte with a differential ability to use bicarbonate. *Freshw. Biol.* 26: 175-187.
- Madsen, T.V. and Sand-Jensen, K. 1991. Photosynthetic carbon assimilation in aquatic macrophytes. *Aquat. Bot.* 41: 5-40.
- Madsen, T.V., Maberly, S.C. and Bowes, G. 1996. Photosynthetic acclimation of submersed angiosperms to CO₂ and HCO₃⁻. *Aquat. Bot.* 53: 15-30.
- Moeller, R.E. 1978. Seasonal changes in biomass, tissue chemistry, and net production of the evergreen hydrophyte, *Lobelia dortmanna*. *Can. J. Bot.* 56: 1425-1433.
- Morton, B.A. and Keeley, J.E. 1990. C₄ Acid fixation in photosynthesis of the submerged aquatic *Eleocharis acicularis* (L.) R. & S. *Aquat. Bot.* 36: 379-388.
- Nielsen, S.L. and Sand-Jensen, K. 1993. Photosynthetic implications of heterophylly in *Batrachium peltatum* (Schränk) Presl. *Aquat. Bot.* 44: 361-371.
- Pedersen, P., Sand-Jensen, K. and Revsbech, N.P. 1995. Diel pulses of O₂ and CO₂ in sandy lake sediments inhabited by *Lobelia dortmanna*. *Ecology* 76 (5): 1536-1545.
- Penuelas, J. 1985. HCO₃⁻ as an exogenous carbon source for aquatic bryophytes *Fontinalis antipyretica* and *Fissidens grandifrons*. *J. Exp. Bot.* 36: 441-448.
- Prins, H.B.A. and Elzenga, J.T.M. 1989. Bicarbonate utilization: function and mechanism. *Aquat. Bot.* 34: 59-83.
- Raven, J.A. 1970. Exogenous inorganic carbon sources in plant photosynthesis. *Biol. Rev.* 45: 167-221.
- Raven, J.A. 1995. Photosynthesis in aquatic plants. In: Schulze, E.D., Caldwell, M. (eds.). *Ecophysiology of photosynthesis*. 299-318.
- Richardson, K., Griffiths, H., Reed, M.L., Raven, J.A. and Griffiths, N.M. 1984. Inorganic carbon assimilation in the Isoetids, *Isoetes lacustris* L. and *Lobelia dortmanna* L. *Oecologia* 61: 115-121.
- Reiskind, J.B., Berg, R.H., Salvucci, M.E. and Bowes, G. 1989. Immunogold localization of primary carboxylases in leaves of aquatic and a C₃-C₄ intermediate species. *Plant Science* 61: 43-52.
- Reiskind, J., Madsen, T.V., van Ginkel, L.C. and Bowes, G. 1997. Evidence that inducible C₄-type photosynthesis is a chloroplastic CO₂-concentrating mechanism in *Hydrilla*, a submerged monocot. *Plant Cell Environ.* 20: 211-220.
- Roelofs, J.G.M., Schuurkes, J.A.A.R. and Smits, A.J.M. 1984. Impact of acidification and eutrophication on macrophyte communities in soft waters. II. Experimental studies. *Aquat. Bot.* 18: 389-411.
- Sand-Jensen, K. and Frost-Christensen, H. 1999. Plant growth and photosynthesis in the transition zone between land and stream. *Aquat. Bot.* 63: 23-35.
- Sand-Jensen, K. and Gordon, D.M. 1993. Variable

- HCO_3^- affinity of *Elodea canadensis* Michaux in response to different HCO_3^- and CO_2 concentration during growth. *Oecologia* 70: 426-432.
- Sculthorpe, C.D. 1967. *The Biology of Aquatic Vascular Plants*. London.
- Smart, R.M. and Barko, J.W. 1985. Laboratory culture of submersed freshwater macrophytes on natural sediments. *Aquat. Bot.* 21: 251-263.
- Spence, D.H.N. and Maberly, S.C. 1985. Occurrence and ecological importance of HCO_3^- use among aquatic higher plants. In: Lucas, W.J., Berry, J.A. (eds.). *Inorganic carbon uptake by aquatic photosynthetic organisms*. 125-143.
- Steemann-Nielsen, E. 1947. Photosynthesis of aquatic plants with special references to the carbon sources. *Dansk Bot. Arkiv* 12: 3-71.
- Stumm, W. and Morgan, J.J. 1996. *Aquatic Chemistry*. New York
- Wium-Andersen, S. 1971. Photosynthetic uptake of free CO_2 by the roots of *Lobelia dortmanna*. *Physiol. Plant.* 25: 245-248.
- Wium-Andersen, S. and Andersen, J.M. 1972. Carbon dioxide content of the interstitial water in the sediment of Grane Langsø, a Danish *Lobelia* lake. *Limnol. Oceanogr.* 17: 943-947.